Vasopressin release by antagonists of GABA and glycine

W. FELDBERG & M. ROCHA E. SILVA JR

National Institute for Medical Research, Mill Hill, London NW7 1AA

In cats anaesthetized with chloralose, nicotine releases vasopressin without oxytocin when acting from the ventral surface of the brain stem. To obtain this effect nicotine was injected into the cerebral ventricles or. more usually, applied topically through perspex rings (20 µl. into each ring of a solution of 2 or 4 mg/ml) to a bilateral region lateral to the pyramids and 6 to 9 mm caudal to the trapezoid bodies (Bisset, Feldberg, Guertzenstein & Rocha e Silva Jr. 1975). The vasopressin release occurred almost instantaneously and was short-lasting. It was followed by a paralyzing action rendering the region insensitive to renewed nicotine application. Hexamethonium similarly applied shared with nicotine the paralyzing action but did not itself stimulate vasopressin release (Bisset & Feldberg, unpublished experiments).

Using the procedure of Bisset et al., there was no vasopressin release on topical application of physostigmine (10 to 50 mg/ml) carbachol (6 mg/ml), tetra-ethyl and tetra-methyl ammonium iodide (5–20 mg/ml) to the nicotine sensitive region, nor on injection into the cerebral ventricles of noradrenaline (100 μ g) or morphine sulphate (750 μ g), the morphine being injected into unanaesthetized cats. But the topical application of tubocurarine (20 mg/ml), for only a few minutes, released large amounts of vasopressin. In contrast to the effect of nicotine the release proceeded gradually and reached its maximum after about 1 hour.

On account of its neuromuscular and ganglion blocking action, tubocurarine might have been expected to act like hexamethonium. However, in the brain, tubocurarine produces strong long-lasting excitation thought to be due, at least partly, to disinhibition, i.e., antagonizing the action of GABA released from central inhibitory neurones. If this were the mechanism of its vasopressin releasing property other GABA antagonists, like picrotoxin, bicuculline and leptazol should have the same effect, and so they had. With regard to their other central actions, picrotoxin and bicuculline were found to be more, and leptazol less potent than tubocurarine; the leptazol effect was also of much shorter duration. The same differences seem to apply to their vasopressin releasing property. On topical application through the perspex rings, picrotoxin and bicuculline were effective in concentrations of 2 mg/ml., leptazol of 50 mg/ml. With leptazol the release ceased shortly after its removal, but continued when left inside the rings. Strychnine, an inhibitor of glycine, also released vasopressin on application to the nicotine sensitive region in concentrations of 5-10 mg/ml.

It is suggested that vasopressin release in the body is continuously inhibited by inhibitory neurones in the medulla which release GABA and glycine.

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Reference

BISSET, G.W., FELDBERG, W., GUERTZENSTEIN, P.G. & ROCHA E SILVA, Jr. M. (1975). Vasopressin release by nicotine: the site of action. *Br. J. Pharmac.*, 54, 463-474.

Morphine selectively blocks dopamine-stimulated cyclic AMP formation in rat neostriatal slices

K.P. MINNEMAN (introduced by L.L. IVERSEN)

MRC Neurochemical Pharmacology Unit, Department of Pharmacology, University of Cambridge

Many of the varied effects of the opiate narcotics may be due to a primary interaction with cyclic nucleotide systems. In both rat brain and cultured cells, morphine markedly affects both cyclic AMP and cyclic GMP metabolism (Minneman & Iversen, 1976). Behavioural and neurochemical evidence suggests that morphine may antagonize the effects of dopamine *in vivo*, in a manner differing significantly from neuroleptic drugs (Kuschinsky & Hornykiewicz, 1972). We report here a potent inhibitory effect of morphine on dopaminestimulated cyclic AMP formation in intact slices of rat neostriatum.

Slices $(260 \times 260 \,\mu\text{m})$ were prepared from rat neostriata as described previously (Minneman & Iversen, 1976). Drugs and preincubated tissue were added to a final volume of 250 μ l medium and incubated for 15 min with no added phosphodiesterase inhibitor. The reaction was stopped by boiling and the